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DATE: Tuesday, July 27, 2004

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☐	L4	(l1 and L2) and (duplex)	5
☐	L5	6420115.pn. or 6403313.pn.	5
☐	L6	L5 and (parallel near$3 complementary)	0
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☐	L8	L5 and (parallel near complementary)	0
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☐	L10	(l1 or L2) same target	29
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☐	L13	(l1 and L2) and (nucleic acid or oligonucleotide or probe or primer)	14

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Database: US Pre-Grant Publication Full-Text Database; US Patents Full-Text Database; US OCR Full-Text Database; EPO Abstracts Database; JPO Abstracts Database; Derwent World Patents Index; IBM Technical Disclosure Bulletins

Search: L9 and oligonucleotide

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DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ			
L1	bolli-M$.in.	30	L1
L2	Huldreich-T$.in. or leumann-C$.in.	6	L2
L3	(l1 or l2) and bicyclo-DNA	0	L3
L4	(l1 or l2) and DNA	3	L4
L5	(l1 or l2) and parallel	3	L5
L6	(l1 or l2) and (parallel same complementary)	0	L6
L7	parallel complementary	513	L7
L8	L7 and (DNA or nucleic acid or oligonucleotide or RNA)	44	L8
L9	claude-H$.in.	166	L9

Double helixes with **parallel** strands are formed by nuclease-resistant oligo-[α]-deoxynucleotides and oligo-[α]-deoxynucleotides covalently linked to an intercalating agent with complementary oligo-[β]-deoxynucleotides

AUTHOR(S): Praseuth, Daniele; Chassignol, Marcel; Takasugi, Masashi; Doan, Trung Le; Thuong, Nguyen T.; Helene, Claude

CORPORATE SOURCE: Lab. Biophys., INSERM, Paris, 75005, Fr.

SOURCE: Journal of Molecular Biology (1987), 196(4), 939-42
CODEN: JMOBAK; ISSN: 0022-2836

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Oligo-α-thymidylates were synthesized and covalently linked to an intercalating agent (an acridine derivative) and(or) to a p-azidophenacyl group. These mols. bind to a complementary oligo-β-deoxynucleotide. A strong stabilization is obtained by covalent attachment of the acridine derivative at the 5' end of the oligo-α-deoxynucleotide. Upon excitation of the p-azidophenacyl group with UV light, the oligo-α-thymidylate is crosslinked to its target sequence. These crosslinks are converted to chain breaks under alkaline conditions. This allows an unambiguous assignment of the orientation of the 2 oligonucleotide chains. As expected, β-β hybrids have an antiparallel orientation, whereas the 2 chains of α-β hybrids are **parallel** independently of whether an intercalating agent is covalently linked to the .**alpha**.-**oligonucleotide**. Oligo-α-thymidylates covalently linked to an acridine derivative are highly resistant to endo- and exonucleases. Therefore, they could be used as anti-messengers to block mRNA translation in vivo under conditions where oligo-β-deoxynucleotides are usually hydrolyzed.

=>

Sequence-specific recognition, photocrosslinking and cleavage of the DNA double helix by an oligo-[α]-thymidylate covalently linked to an azidoproflavine derivative

AUTHOR(S): Trung Le Doan; Perrouault, Loic; Praseuth, Daniele; Habhoub, Noureddine; Decout, Jean Luc; Nguyen Thanh Thuong; Lhomme, Jean; Helene, Claude

CORPORATE SOURCE: Lab. Biophys., Museum Natl. Hist. Nat., Paris, 75005, Fr.

SOURCE: Nucleic Acids Research (1987), 15(19), 7749-60
CODEN: NARHAD; ISSN: 0305-1048

DOCUMENT TYPE: Journal

LANGUAGE: English

AB A 3-azidoproflavine derivative was covalently linked to the 5'-end of an octathymidylate synthesized with the α-anomers of the nucleoside. Two target nucleic acids were used for this substituted oligo(α-thymidylate): a 27-mer single-stranded DNA fragment containing an octadeoxyadenylate sequence and a 27-mer duplex containing 8 contiguous A·T base pairs with all adenines on the same strand. Upon visible light irradiation the octa-[α]-thymidylate was photocrosslinked to the single-stranded 27-mer. Chain breaks were induced at the crosslinked sites upon piperidine treatment. From the location of the cleavage sites on the 27-mer sequence it was concluded that a triple helix was formed by the azidoproflavine-substituted oligo-[α]-thymidylate with its complementary oligodeoxyadenylate sequence. When the 27-mer duplex was used as a substrate, cleavage sites were observed on both strands after piperidine treatment of the irradiated sample. They were located at well defined positions which indicated that the octathymidylate was bound to the (dA)8·(dT)8 sequence in a **parallel** orientation with respect to the (dA)8-containing strand. Specific binding of the [α]-octathymidylate involved local triple strand formation with the duplex (dA)8·(dT)8 sequence. Thus, it is possible to synthesize sequence-specific mols. which specifically bind oligopurine-oligopyrimidine sequences in double-stranded DNA via recognition of the major groove H bonding sites of the purines.

mparative activity of alpha- and beta-anomeric oligonucleotides on rabbit beta globin synthesis: inhibitory effect of cap targeted **alpha-oligonucleotides**.

COMMENT: Erratum in: Biochem Biophys Res Commun 1989 Dec 29;165(3):1443

AUTHOR: Bertrand J R; Rayner B; Imbach J L; Paoletti C; Malvy C

CORPORATE SOURCE: UA 147 CNRS, U 140 INSERM, Institut Gustave Roussy, Villejuif, France.

SOURCE: Biochemical and biophysical research communications, (1989 Oct 16) 164 (1) 311-8.
Journal code: 0372516. ISSN: 0006-291X.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 198911

ENTRY DATE: Entered STN: 19900328
Last Updated on STN: 19900328
Entered Medline: 19891120

AB Alpha-anomeric oligonucleotides are resistant to nucleases and display **parallel** annealing to RNA complementary sequences. We compared the effect of alpha- and beta-oligonucleotides targeted against various mRNA regions on the rabbit beta globin in vitro synthesis. In order to determine the role of RNase H, experiments were performed in both rabbit reticulocyte lysate and wheat germ extract. As expected beta-oligonucleotides were found more efficient in wheat germ extract which is rich in RNase H activity and **alpha-oligonucleotide** targeted against the initiation codon or downstream had no effect because they do not induce mRNA cleavage by RNase H. However, we report, for the first time, a specific translation inhibition by **alpha-oligonucleotides**. This occurs provided they are targeted against the cap region in 5' of the mRNA.

PubMed ID: 9461474

TITLE: alpha-Oligodeoxyribonucleotide N3'-->P5' phosphoramidates: synthesis and duplex formation.

AUTHOR: Pongracz K; Gryaznov S M

CORPORATE SOURCE: Lynx Therapeutics Inc., 3832 Bay Center Place, Hayward, CA 94545, USA.

CONTRACT NUMBER: RR01614 (NCRR)

SOURCE: Nucleic acids research, (1998 Feb 15) 26 (4) 1099-106. Journal code: 0411011. ISSN: 0305-1048.

PUB. COUNTRY: ENGLAND: United Kingdom

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199803

ENTRY DATE: Entered STN: 19980326
Last Updated on STN: 19980326
Entered Medline: 19980317

AB The synthesis and hybridization properties of novel nucleic acid analogs, alpha-anomeric oligodeoxyribonucleotide N3'-->P5' phosphoramidates, are described. The alpha-3'-aminonucleoside building blocks used for oligonucleotide synthesis were synthesized from 3'-azido-3'-deoxythymidine or 3'-azido-2',3'-dideoxyuridine via acid catalyzed anomerization or transglycosylation reactions. The base-protected alpha-5'-O-DMT-3'-aminonucleosides were assembled into dimers and oligonucleotides on a solid support using the oxidative phosphorylation method.1H NMR analysis of the alpha-N3'-->P5' phosphoramidate dimer structures indicates significant differences in the sugar puckering of these compounds relative to the beta-N3'-->P5' phosphoramidates and to the alpha-phosphodiester counterparts. Additionally, the ability of the **alpha-oligonucleotide** N3'-->P5' phosphoramidates to form duplexes was studied using thermal denaturation experiments. Thus the N3'-->P5' phosphoramidate decamer containing only alpha-thymidine residues did not bind to poly(A) and exhibited lower duplex thermal stability with poly(dA) than that for the corresponding beta-anomeric phosphoramidate counterpart. A mixed base decamer alpha-CTTCTTCCTT formed duplexes with the RNA and DNA complementary strands only in a **parallel** orientation. Melting temperatures of these complexes were significantly lower, by 34-47 or 15-25 degrees C, than for the duplexes formed by the isosequential beta-phosphoramidates in antiparallel and **parallel** orientations respectively. In contrast, the alpha-decaadenylic N3'-->P5' phosphoramidate formed duplexes with both RNA and DNA complementary strands with a stability similar to that of the corresponding beta-anomeric phosphoramidate. Moreover, the self-complementary oligonucleotide alpha-ATATATATAT did not form an alpha:alpha homoduplex. These results demonstrate the effects of 3'-aminonucleoside anomeric configuration on sugar puckering and consequently on stability of the duplexes.

Alpha-DNA.IX: **Parallel** annealing of alpha-anomeric oligodeoxyribonucleotides to natural mRNA is required for interference in RNase H mediated hydrolysis and reverse transcription.

AUTHOR: Gagnor C; Rayner B; Leonetti J P; Imbach J L; Lebleu B
CORPORATE SOURCE: Laboratoire de Biochimie des Proteines, UA 1191 CNRS, Universite des Sciences et Techniques du Languedoc, Montpellier, France.

SOURCE: Nucleic acids research, (1989 Jul 11) 17 (13) 5107-14. Journal code: 0411011. ISSN: 0305-1048.
PUB. COUNTRY: ENGLAND: United Kingdom
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 198909
ENTRY DATE: Entered STN: 19900309
Last Updated on STN: 19960129
Entered Medline: 19890911

AB ps- and aps-alpha anomeric oligodeoxyribonucleotides were designed to recognize in **parallel** (ps) or antiparallel (aps) orientation two different sites of a 1000 base-long mRNA. Northern blots experiments indicate that only ps-**alpha-oligonucleotides** were able to hybridize to the mRNA target. Furthermore, only ps-**alpha-oligonucleotides** were able, in a sequence specific way, to protect the mRNA target against RNase H mediated hydrolysis or to inactivate the priming capacity of beta-oligodeoxynucleotide probes in reverse transcription. Formation of **parallel**-stranded mRNA **alpha-oligonucleotide** miniduplexes which prevents hybridization of beta-oligonucleotide probes is the most likely mechanism accounting for these results. Use of **alpha-oligonucleotides** as potential gene control agents is discussed.

L6 ANSWER 8 OF 12 MEDLINE on STN

Inhibition of Moloney murine leukemia virus reverse transcriptase by alpha-anomeric oligonucleotides.

AUTHOR: Lavignon M; Bertrand J R; Rayner B; Imbach J L; Malvy C; Paoletti C

CORPORATE SOURCE: UA 147 CNRS, U 140 INSERM, Institut Gustave Roussy, Villejuif, France.

SOURCE: Biochemical and biophysical research communications, (1989 Jun 30) 161 (3) 1184-90.
Journal code: 0372516. ISSN: 0006-291X.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 198908

ENTRY DATE: Entered STN: 19900309
Last Updated on STN: 19970203
Entered Medline: 19890808

AB After **parallel** hybridization to complementary template RNA, alpha-anomeric oligonucleotides are not primers for Moloney murine leukemia virus reverse transcriptase. As can be expected they are competitors of classical primer oligonucleotides (beta-anomeric). They therefore inhibit the RNA dependent DNA polymerase activity of Moloney murine leukemia virus reverse transcriptase with either homopolymeric or heteropolymeric substrates. Non complementary **alpha-oligonucleotides** display no inhibitory activity. **alpha-Oligonucleotides** are therefore potential candidates for inhibition of retroviral reverse transcriptases by interference with the primer binding sites.

Monoclonal antibodies targeted to **alpha-oligonucleotides**. Characterisation and application in nucleic acid detection.

AUTHOR: Cros P; Kurfurst R; Allibert P; Battail N; Piga N; Roig V; Thuong N T; Mandrand B; Helene C

CORPORATE SOURCE: Laboratoire des Sondes Nucleiques, bioMerieux, ENS, Lyon, France.

SOURCE: Nucleic acids research, (1994 Aug 11) 22 (15) 2951-7. Journal code: 0411011. ISSN: 0305-1048.

PUB. COUNTRY: ENGLAND: United Kingdom

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199409

ENTRY DATE: Entered STN: 19941005
Last Updated on STN: 19960129
Entered Medline: 19940921

AB The aim of the present study was to test the antigenicity of alpha-deoxyribonucleotides in order to develop a new tool for the detection of nucleic acid sequences for use in diagnostic applications. We describe four monoclonal antibodies (Mabs) which recognize alpha-deoxyribonucleotides. Two were raised against a poly(alpha-dT) sequence and specifically recognized the alpha-dT nucleotide. Two were raised against a sequence containing all four common nucleotides as alpha-nucleotides and, surprisingly, only recognized the alpha-dG nucleotide. For all four Mabs, no cross reactivity was observed with beta-oligonucleotides. These Mabs were reactive with **alpha-oligonucleotide** sequences whether these sequences were single-stranded or hybridized to DNA or RNA. The four Mabs were tested in a sandwich hybridization assay that consisted of an **alpha-oligonucleotide** (for target sequence recognition), one of the four Mabs (for recognition of the hybridized **alpha-oligonucleotide**), and goat anti-mouse antibody conjugated to horse radish peroxidase (HRP) (for detection). One of the monoclonal antibodies, Mab 2E11D7, was directly conjugated to HRP and used in sandwich hybridization to detect **PCR** fragments of HPV 18 DNA. The sensitivity of this reaction was 1 pg of plasmid DNA containing the HPV 18 fragment. The specificity of the detection was demonstrated using HPV 6/11 and 16 DNA sequences.

ACCESSION NUMBER: 92346248 EMBASE

DOCUMENT NUMBER: 1992346248

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